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# 50

## Cephalopoda

Bernd U. Budelmann

*Note: This short chapter has not been written to stand on its own. It is an update of Peter Boyle's Chapter 7 in the 7th edition of the UFAW Handbook, which was published in 1999 and is reprinted after this update. Consequently, for details and earlier references the reader should consult that chapter as the authoritative text. In addition, some key literature on cephalopods published earlier (but not mentioned in that chapter) has been added. For an easy follow-up and comparison, the updates given below use (sub)headings similar to those of the earlier chapter. This chapter update is dedicated to the memory of Peter Boyle.*

### Introduction

Cephalopods are the only invertebrate animal group included in this edition of the Handbook, however information on decapod crustaceans is available in the 7th edition of the Handbook. Invertebrates are not generally protected in animal welfare legislation, although certain species are included in some national legislation (eg, UK, New Zealand & Australian Capital Territories, and some Scandinavian countries). The UK, at the time of writing, includes one species, *Octopus vulgaris*, in its legislation on the use of animals in research. However, there is debate within Europe (unresolved at the time of writing) as to whether some, all or no cephalopods (as well as decapod crustacea) should be included in a new European Directive on research using animals (see also Chapter 8 on legislation).

Cephalopods belong to the phylum Mollusca and thus are close relatives to gastropods and bivalves. Although the cephalopod body design conserved some typical molluscan features, it developed a level of complexity, and especially a sophisticated nervous system and sense organs, that in several aspects reach vertebrate standards (Budelmann 1995). The cephalopod nervous system is certainly the most advanced of any invertebrate nervous system and this complexity correlates well with the animals' (in general) very active, fast moving predatory life styles and sophisticated behaviours (Bullock and Horridge 1965; Budelmann 1995; Hanlon & Messenger 1996). Not surprisingly then, beginning in the 1930s with J.Z. Young's rediscovery of the giant axon and the subsequent early understanding of the processes involved in nerve impulse conduction and transmission (Adelman & Gilbert 1990), cephalopods have become fascinating and valuable invertebrate model systems for comparative vertebrate research. However, in such comparisons a number of limitations apply (see below, nervous

system). It is important that these limitations should be understood because of the recent increased interest in this animal group from media and laymen that is sometimes combined with a tendency for 'over-interpretation' of the fascinating behaviours that cephalopods show.

The class Cephalopoda comprises two sub-classes: Nautiloidea (*Nautilus*) and Coleoidea (octopuses, cuttlefish and squids). The latter two are often referred to as 'decapods' but care should be taken as this term is also commonly used for the order of crustaceans that includes crabs, lobsters, prawns, etc). Nautiloids and coleoids show significant differences with regard to their anatomy, physiology and behaviour (for example, nautilus have a much less sophisticated nervous system and sensory outfit; they show no colour change and have a much simpler behavioural repertoire; and they are scavengers and thus seem to have a well developed chemosensory system for food detection). These differences between nautiloids and coleoids, however, unfortunately are often not considered in the literature when the term 'cephalopod(s)' is applied. The term as used often refers to the coleoid cephalopods only (that is, to the octopuses, cuttlefish and squids) and, therefore, great care must be taken in the use, interpretation and generalisation of the 'cephalopod' data described.

### Cephalopod biology

A number of comprehensive monographs has become available that cover almost all aspects of cephalopod biology, from palaeobiology (Landman *et al.* 1996, 2007), evolution, systematics, identification, and biogeography (Clarke 1986; Guerra 1992; Sweeney *et al.* 1992; Payne *et al.* 1998; Voss *et al.* 1998; Norman 2000; Capua 2004; Jereb & Roper 2005), to gross and microscopic anatomy (Mangold 1989; Budelmann *et al.* 1997; Nixon & Young 2003), physiology (Abbott *et al.* 1995), ecology, fisheries and culture (Boucaud-Camou 1991; Boyle & Rodhouse 2005; Chotiyaputta *et al.* 2005), age determination (Jereb *et al.* 1991), behaviour (Hanlon & Messenger 1996; Nixon & Young 2003; Borelli & Fiorito 2008) and diseases (Hanlon & Forsythe 1990; Hochberg 1990). Some more recent data are summarised below.

### Habitat and distribution

An excellent monograph is now available on many aspects of cephalopod ecology and fisheries, including: cephalopod



biodiversity and zoogeography; life cycle, growth and reproduction; population ecology; cephalopods as prey and predators; fishing methods and scientific sampling; fishery resources; fisheries oceanography; and assessment and management (Boyle & Rodhouse 2005).

## Locomotion

The cephalopod musculature lacks a skeletal support system and, instead, operates on the principle of a muscular hydrostat, similar to the 'mechanism' of an elephant trunk or the human tongue (Kier & Smith 1985; Smith & Kier 1989). This allows cephalopod (specifically octopod) arms a great range of movement. Over the past more than 10 years, significant progress has been made in understanding the nervous control of cephalopod arm movements; this ultimately could inspire completely new strategies for the control of highly flexible robotic arms (Gutfreund *et al.* 1996, 1998; Matzner *et al.* 2000; Sumbre *et al.* 2001, 2005; Walker *et al.* 2005; Yekutieli *et al.* 2005a, 2005b, 2007).

## Shell and buoyancy

The neutral buoyancy of many squids is achieved by storing ammonia in body tissue. The various mechanisms for storage (in coelomic cavities, vacuoles, or gelatinous outer layers) have been reviewed in support for the argument that ammoniacal squids have evolved as a polyphyletic animal group (Voight *et al.* 1994).

## Respiration and circulation

Water temperature, pH and oxygen supply are critical factors in cephalopod culture and breeding. Recently, the cuttlefish (*Sepia officinalis*) has served as a valuable model system for understanding the mechanisms of thermal tolerance in ectothermic animals (Melzner *et al.* 2006, 2007).

## Nervous system

The cephalopod nervous system is the most highly evolved of all invertebrate nervous systems. On the other hand and despite that level of complexity, the overall organisation of its central part (the brain) is fundamentally different from that of the vertebrate nervous system and, therefore, any direct comparison between the two has serious limitations. This, however, neither excludes careful comparison of basic brain functions, nor weakens the great value of the cephalopod nervous system in comparative research. Details of the anatomy of the cephalopod central nervous system are available for *Nautilus* (Young 1965), *Octopus* (Young 1971; Budelmann & Young 1985; Plän 1987) and *Loligo* (Young 1974, 1976, 1977, 1979; Messenger 1979; Budelmann & Young 1987); recent overviews are given in Budelmann (1995), Budelmann *et al.* (1997), Nixon and Young (2003) and Williamson and Chrachri (2004).

In addition to their highest level of complexity, cephalopod brains are also the largest of all the invertebrate brains; their brain:body weight ratio exceeds that of many fishes and reptiles (Packard 1972). This is not too surprising, however, since cephalopods lack an internal skeleton and lack joints, and thus lack a 'simple' antagonistic muscle control of movements. Consequently, about half of the volume of the brain of coleoid cephalopods consists of the relatively large motoneurons that form the sub-oesophageal mass of the brain; this area includes the motoneurons that expand all the chromatophore organs in the skin (for a summary of the numbers of nerve cells in the various parts of the *Octopus* brain, see Budelmann 1995). On the other hand, the comparatively large size and complexity of the brains of octopuses, cuttlefish and squids are the basis for the animals' large repertoire of fascinating behaviours, including various forms of learning and short- and long-term memory (Hanlon & Messenger 1996; Hochner *et al.* 2003; Borelli & Fiorito 2008). Ultimately, these make coleoid cephalopods, especially shallow-water octopods, cuttlefish and squids, the only invertebrate species with which humans can directly interact ('communicate') in a back and forth manner and beyond a simple reflex-like (re)action on the animals' side.

Octopuses, cuttlefish and squids are often considered the most 'intelligent' invertebrate species (whichever way intelligence is defined). The web-based Wikipedia summarises this issue quite well:

*Cephalopod intelligence has an important comparative aspect in our understanding of intelligence, because it relies on a nervous system fundamentally different from that of vertebrates. ... The scope of cephalopod intelligence is controversial ... Classical conditioning of cephalopods has been reported, and one study (Fiorito and Scotto 1992) even concluded that octopuses practice observational learning. However, the latter idea is strongly disputed, and doubt has been shed on some other reported capabilities as well. In any case, impressive spatial learning capacity, navigational abilities, and predatory techniques remain beyond question.*

Other impressive behaviours that can be added to this list are cephalopod mating and social behaviours, including social recognition (Hanlon & Messenger 1996; Boal *et al.* 2000; Dickel *et al.* 2000; Karson *et al.* 2003; Boal 2006; Alves *et al.* 2008; Borelli & Fiorito 2008).

Powerful techniques have now successfully been applied to cephalopod brains to the study of their anatomy and function (Budelmann *et al.* 1995): three-dimensional magnetic resonance imaging of brain pathways (Quast *et al.* 2001) and individual neurons (Gozansky *et al.* 2003); brain slice recordings (Williamson & Budelmann 1991); recordings from intact animals with implanted electrodes (Bullock & Budelmann 1991); and mapping of metabolic brain activity (Novicki *et al.* 1992). With these modern neurophysiological techniques cephalopods have become an increasingly valuable invertebrate model system for comparative vertebrate research, such as the evolution of learning and memory and other higher brain functions (Hochner *et al.* 2006). Recently, laterality in the brain (Byrne *et al.* 2002, 2004, 2006), play behaviour (Kuba *et al.* 2006), personality (Sinn & Moltschanowskyj 2005), sleep (Brown *et al.* 2006) and



complex phenomena, such as consciousness and suffering in cephalopods, have been discussed (Mather 2001, 2008).

## Sense organs

Cephalopods have a sophisticated sensory outfit that includes all major sense organs, such as photoreceptors (including extra-ocular photoreceptors), distance and contact chemoreceptors, and various mechanoreceptors (including equilibrium receptor organs, a lateral line analogue system and a neck proprioceptive organ) (Budelman 1996). Knowledge about touch, pressure and muscle proprioceptors is limited, and it still remains to be seen whether cephalopods have electroreceptors and are sensitive to pain. For a comprehensive summary on cephalopod sense organs, see Budelman *et al.* (1997).

## Eyes and vision

The cephalopod and vertebrate lens eyes are a textbook example of analogy (convergent evolution) between an invertebrate and a vertebrate sensory system. Recent advances have been made in understanding visual processing (Chrachri & Williamson 2003, 2004, 2005; Chrachri *et al.* 2005; Douglas *et al.* 2005), and the role of polarised vision (Saidel *et al.* 1983; Shashar *et al.* 1998, 2002; Boal *et al.* 2004; Saidel *et al.* 2005; Mäthger & Hanlon 2006). In addition, in the squid *Lolliguncula* a dorsal light reflex has been described (Preuss & Budelman 1995a) and in cuttlefish a counter-shading reflex (Ferguson *et al.* 1994).

## Equilibrium receptor organs

A tremendous body of data has accumulated over the past 40 years on the anatomy, ultrastructure and physiology of the cephalopod equilibrium receptor organs (statocysts), which include sophisticated receptor systems for the detection of linear (gravity) and angular accelerations (Budelman *et al.* 1987; for summaries, see Budelman 1990; Budelman *et al.* 1997; for the *Nautilus* statocyst, see Neumeister & Budelman 1997). Special emphasis has been paid to the similarities between the structure and function of the cephalopod and vertebrate hair cells (eg, Budelman & Williamson 1994; Budelman 2000), including their ion channels and efferent innervation (Williamson 1995) and transmitter and transmitter-like substances; the latter include nitric oxide (Tu & Budelman 2000) and cannabinoids (Tu & Budelman, unpublished). Cephalopod statocysts are known to drive a sophisticated control system for compensatory eye movements; part of its central organisation resembles that of the vertebrate vestibulo-oculomotor reflex pathway and involves four (*Nautilus*), seven (octopods), or 13–14 (cuttlefish and squids) extra-ocular eye muscles (Budelman & Young 1984, 1993; Neumeister & Budelman 1997).

## Epidermal lines

The epidermal lines (formerly known as 'Drüsenlinien') that occur on the head and arms of at least some of the coleoid species are analogous to the fish and aquatic amphibian

lateral line systems (Budelman & Bleckmann 1988; Budelman *et al.* 1997).

## Neck proprioceptive organ

Similar to the vertebrate neck muscle proprioceptors, cuttlefish and squid have groups of epidermal hair cells on their neck that serve as a proprioceptive neck organ for the control of the position of the head relative to the body (Preuss & Budelman 1995b).

## Vibration receptors and hearing

Cephalopods are sensitive to vibrational stimuli via statocyst receptors and sense local water movements with their lateral line analogue system (Budelman & Bleckmann 1988; Williamson 1988; Packard *et al.* 1990; Bleckmann *et al.* 1991; Komak *et al.* 2005). On the other hand, there is much confusion regarding cephalopods ability to 'hear'. Ultimately, this is a semantic issue since the answer depends on the definition of underwater sound and underwater hearing (Budelman 1992). In conventional terms, cephalopods cannot hear because they do not have a receptor system that is specialised for the detection of the pressure wave of underwater sound.

## Maintenance, culture and laboratory procedures

With the growing interest in cephalopods for research and commercial mariculture, as well as their popularity in public and private displays, knowledge about the maintenance, culture and proper laboratory procedures is of increased importance (Oestmann *et al.* 1997; Sykes *et al.* 2006; Dunlop & King 2008). For advice on optimising the survival of hatchling cuttlefish and squid, see Forsythe *et al.* (1994), Vidal *et al.* (2002a, 2002b) and Sykes *et al.* (2003). With regard to culture density, recent cuttlefish data show that lower stocking density results in better growth (Domingues *et al.* 2003; Correia *et al.* 2005), as does higher water temperature (25°C compared to 17°C; Forsythe *et al.* 2002). On the other hand, lower temperature (15°C compared to 27°C) extends the life cycle (Domingues *et al.* 2002). For the development of memory in cuttlefish, an enriched environment is crucial during their second and/or third months of life (Dickel *et al.* 2000). Crowding of adult cuttlefish should be avoided since it stimulates aggression (Boal *et al.* 1999).

Not surprisingly, the quality and composition of food has been proven to be critical for good growth and survival. Adult cuttlefish show much better growth rates when fed with live or thawed natural prey than when fed with an artificial diet (Domingues *et al.* 2005). Shrimp-based food pellets, although less palatable, produce maintenance growth, whereas a highly palatable fish-based surimi diet (mimicking the meat of lobster, crab and other shellfish) results in poor survival (Castro *et al.* 1993). When fed with live mysid shrimp, grass shrimp, or fish fry, young cuttlefish showed best growth during the first week after hatching when fed with mysid shrimp, and thereafter when fed with grass shrimp; cuttlefish fed with fish fry showed lowest growth rates at all times (Domingues *et al.* 2004). When prey



is maintained for feeding juvenile cuttlefish, prey starvation should be avoided (Correia *et al.* 2009). Although *Sepia* shows an innate food preference, early familiarisation with other food can override this preference (Darmaillacq *et al.* 2006). For the importance of certain elements (including copper and strontium) in the food of octopus, cuttlefish and squid, see Koueta *et al.* (2002) Villanueva and Bustamente (2006) and Iglesias *et al.* (2007).

A comprehensive monograph is now available on invertebrate medicine that includes a chapter on cephalopods (Lewbart 2006) and ethical and welfare considerations for working with cephalopods have recently been summarised (Mather & Anderson 2007; Moltschaniwskyj *et al.* 2007).

## Occupational health hazards

Many octopus and, particularly, cuttlefish and squid species may bite, specifically when stressed, disturbed or improperly handled. Their sharp, parrot-like beaks can inflict significant wounds and the saliva can have a variety of toxic effects. The tetrodotoxin-like venom of the Australian blue-ringed octopus (*Hapalochlaena maculosa*) can be lethal to humans (Williamson *et al.* 1996, for a review).

## Cannibalism

Cannibalism is well known in octopuses, cuttlefish and squids when held in captivity. Obvious reasons include a too high stocking density and an inadequate amount of food and shelter. Specifically, when food supply is limited and feeding *ad libitum* becomes a problem, larger animals may prey upon smaller ones when kept in the same tank. In addition, sexual cannibalism has been described in an octopus species on a coral reef (Hanlon & Forsythe 2008).

## Autophagy

Some data (other than anecdotal) are now available on autophagy (self eating) in *Octopus vulgaris*. They suggest that it is caused by either a substance that is released by the animals themselves or, more likely, by viruses or bacteria; stress may contribute to this behaviour but does not seem to be its primary cause (Budelman 1998).

## Further information and reading

A large number of references to the literature on cephalopods are available from the library service of the Smithsonian Institution Research Information System at <http://sirismm.si.edu/siris/siris-cephalopod.htm>.

The following web pages provide very useful information regarding all major aspects of cephalopod biology, supply and maintenance, rearing, culture and breeding and laboratory procedures (with regard to the scientific accuracy, however, general caution must be taken because of its lack of peer review):

Tree of Life – Cephalopods: <http://tolweb.org/cephalopoda>

Association of Zoos & Aquariums: <http://www.aza.org>

The Cephalopod Page: <http://www.thecephalopodpage.org>

The National Resource Center for Cephalopods: <http://www.cephalopod.org>

Cephbase: <http://www.cephbase.com>

The Octopus News Magazine Online: <http://www.tonmo.com>

## References

- Abbott, N.J., Williamson, R. and Maddock, L. (Eds) (1995) *Cephalopod Neurobiology*. Oxford University Press, Oxford
- Adelman, W.J. and Gilbert, D.L. (1990) Electrophysiology and biophysics of the squid giant axon. In: *Squid as Experimental Animals*. Eds Gilbert, D., Adelman, H. and Arnold, J., pp. 93–132. Plenum Press, New York
- Alves, C., Boal, J.G. and Dickel, L. (2008) Short distance navigation in cephalopods: a review. *Cognitive Processing*, **9**, 239–247
- Bleckmann, H., Budelmann, B.U. and Bullock, T.H. (1991) Peripheral and central nervous responses evoked by small water movements in a cephalopod. *Journal of Comparative Physiology A*, **168**, 247–257
- Boal, J.G. (2006) Social recognition: a top down view of cephalopod behavior. *Vie et Milieu*, **56**, 69–79
- Boal, J.G., Hylton, R.A., Gonzalez, S.A. *et al.* (1999) Effects of crowding on the social behavior of cuttlefish (*Sepia officinalis*). *Contemporary Topics in Laboratory Animal Science*, **38**, 49–55
- Boal, J.G., Dunham, A., Williams, K. *et al.* (2000) Experimental evidence for spatial learning in octopuses. *Journal of Comparative Psychology*, **114**, 246–252
- Boal, J.G., Shashar, N., Grable, M. *et al.* (2004) Behavioral evidence for intraspecific signals with achromatic and polarized light by cuttlefish (Mollusca: Cephalopoda). *Behaviour*, **141**, 837–861
- Borelli, L. and Fiorito, G. (2008) Behavioral analysis of learning and memory in cephalopods. In: *Learning and Memory: A Comprehensive Reference* (Ed. Byrne, J.H.), Vol. I. Ed. Menzel, R., pp. 605–627. Elsevier, Amsterdam
- Boucaud-Camou, E. (Ed.) (1991) *The Cuttlefish*. Centre de Publications de l'Université de Caen, Caen
- Boyle, P. and Rodhouse, P. (2005) *Cephalopods: Ecology and Fisheries*. Blackwell Publishing, Oxford
- Brown, E.R., Piscopo, S., De Stefano, R. *et al.* (2006) Brain and behavioural evidence for rest-activity cycles in *Octopus vulgaris*. *Behavioural Brain Research*, **172**, 355–359
- Budelmann, B.U. (1990) The statocysts of squid. In: *Squid as Experimental Animals*. Eds Gilbert, D., Adelman, H. and Arnold, J., pp. 421–439. Plenum Press, New York
- Budelmann, B.U. (1992) Hearing in non-arthropod invertebrates. In: *The Evolutionary Biology of Hearing*. Eds Webster, B., Fay, R.R. and Popper, A.N., pp. 141–155. Springer, New York
- Budelmann, B.U. (1995) The cephalopod nervous system: What evolution has made of the molluscan design. In: *The Nervous System of Invertebrates: An Evolutionary and Comparative Approach*. Eds Breidbach, O. and Kutsch, W., pp. 115–138. Birkhäuser Verlag, Basel
- Budelmann, B.U. (1996) Active marine predators: The sensory world of cephalopods. *Marine and Freshwater Behavior and Physiology*, **27**, 59–75
- Budelmann, B.U. (1998) Autophagy in *Octopus*. *South African Journal of Marine Science*, **20**, 101–108



- Budelmann, B.U. (2000) Kinociliary mechanoreceptors in the equilibrium receptor organs of cephalopods. In: *Cell and Molecular Biology of the Ear*. Ed. Lim, D.J., pp. 3–17. Kluwer/Plenum Press, New York
- Budelmann, B.U. and Bleckmann, H. (1988) A lateral line analogue in cephalopods: water waves generate microphonic potentials in the epidermal head lines of *Sepia* and *Loliguncula*. *Journal of Comparative Physiology A*, **164**, 1–5
- Budelmann, B.U. and Williamson, R. (1994) Directional sensitivity of hair cell afferents in the *Octopus* statocyst. *Journal of Experimental Biology*, **187**, 245–259
- Budelmann, B.U. and Young, J.Z. (1984) The statocyst-oculomotor system of *Octopus vulgaris*: Eye muscles, eye muscle nerves, statocyst nerves, and the oculomotor centre in the central nervous system. *Philosophical Transactions of the Royal Society London B*, **306**, 159–189
- Budelmann, B.U. and Young, J.Z. (1985) Central pathways of the nerves of the arms and mantle of *Octopus*. *Philosophical Transactions of the Royal Society London B*, **310**, 109–122
- Budelmann, B.U. and Young, J.Z. (1987) Brain pathways of the brachial nerves of *Sepia* and *Loligo*. *Philosophical Transactions of the Royal Society London B*, **315**, 345–352
- Budelmann, B.U. and Young, J.Z. (1993) The oculomotor system of decapod cephalopods: eye muscles, eye muscle nerves, and the oculomotor neurons in the central nervous system. *Philosophical Transactions of the Royal Society London B*, **340**, 93–125
- Budelmann, B.U., Bullock, T.H. and Williamson, R. (1995) Cephalopod brains: promising preparations for brain physiology. In: *Cephalopod Neurobiology*. Eds Abbott, N.J., Williamson, R. and Maddock, L., pp. 399–413. Oxford University Press, Oxford
- Budelmann, B.U., Sachse, M. and Staudigl, M. (1987) The angular acceleration receptor system of *Octopus vulgaris*: morphometry, ultrastructure, and neuronal and synaptic organization. *Philosophical Transactions of the Royal Society London B*, **315**, 305–343
- Budelmann, B.U., Schipp, R. and von Boletzky, S. (1997) Cephalopoda. In: *Microscopic Anatomy of Invertebrates*, Vol. 6A, *Mollusca II*, Eds Harrison, F.W. and Kohn, A., pp. 119–414. Wiley-Liss, New York
- Bullock, T.H. and Budelmann, B.U. (1991) Sensory evoked potentials in unanesthetized unrestrained cuttlefish: a new preparation for brain physiology in cephalopods. *Journal of Comparative Physiology A*, **168**, 141–150
- Bullock, T.H. and Horridge, G.A. (1965) *Structure and Function of the Nervous Systems of Invertebrates*. Freeman, San Francisco
- Byrne, R.A., Kuba, M.J. and Griebel, U. (2002) Lateral asymmetry of eye use in *Octopus vulgaris*. *Animal Behaviour*, **64**, 461–468
- Byrne, R.A., Kuba, M.J. and Meisel, D.V. (2004) Lateralized eye use in *Octopus vulgaris* shows antisymmetrical distribution. *Animal Behaviour*, **68**, 1107–1114
- Byrne, R.A., Kuba, M., Meisel, D.V. et al. (2006) Does *Octopus vulgaris* have preferred arms? *Journal of Comparative Psychology*, **120**, 198–204
- Castro, B.G., DiMarco, F.P., DeRusha, R.H. et al. (1993) The effects of surimi and pelleted diets on the laboratory survival, growth and feeding rate of the cuttlefish *Sepia officinalis* L. *Journal of Experimental Marine Biology and Ecology*, **170**, 241–252
- Capua, D. (2004) I Cefalopodi dell coste e dell 'arcipelago Toscano: Sistematica, anatomia, fisiologia e sfruttamento delle specie presenti nel Mediterraneo. ConchBooks, Hackenheim
- Chotiyaputta, C., Hatfield, E.M.C. and Lu, C.C. (eds) (2005) *Cephalopod Biology, Recruitment and Culture*. Phuket Marine Biological Center Research Bulletin No. 66, Phuket
- Chrachri, A. and Williamson, R. (2003) Modulation of spontaneous and evoked EPSCs and IPSCs in optic lobe neurons of cuttlefish *Sepia officinalis* by the neuropeptide FMRF-amide. *European Journal of Neuroscience*, **17**, 1–11
- Chrachri, A. and Williamson, R. (2004) Cholinergic and glutamatergic spontaneous and evoked excitatory postsynaptic currents in optic lobe neurons of cuttlefish, *Sepia officinalis*. *Brain Research*, **1020**, 178–187
- Chrachri, A. and Williamson, R. (2005) Dopamin modulates synaptic activity in the optic lobes of cuttlefish, *Sepia officinalis*. *Neuroscience Letters*, **377**, 152–157
- Chrachri, A., Nelson, L. and Williamson, R. (2005) Whole-cell recording of light-evoked photoreceptor responses in a slice preparation of the cuttlefish retina. *Visual Neuroscience*, **22**, 359–370
- Clarke, M.R. (Ed.) (1986) *A Handbook for the Identification of Cephalopod Beaks*. Clarendon Press, Oxford
- Correia, M., Domingues, P.M., Sykes, A. et al. (2005) Effects of culture density on growth and broodstock management of the cuttlefish, *Sepia officinalis* (Linnaeus, 1758). *Aquaculture*, **245**, 163–173.
- Correia, M., Palma, J., Kirakowski, T. and Andrade J.P. (2009) Effects of prey nutritional quality on the growth and survival of juvenile cuttlefish, *Sepia officinalis* (Linnaeus, 1758). *Aquaculture Research*, **39**, 869–876
- Darmaillacq, A.S., Chichery, R., Shashar, N. and Dickel, L. (2006) Early familiarization overrides innate prey preference in newly hatched *Sepia officinalis* cuttlefish. *Animal Behaviour*, **71**, 511–514
- Dickel, L., Boal, J.G. and Budelmann, B.U. (2000) Effect of early experience on learning and memory in cuttlefish. *Developmental Psychobiology*, **36**, 101–110
- Domingues, P.M., Sykes, A. and Andrade, J.P. (2002) The effects of temperature in the life cycle of two consecutive generations of the cuttlefish *Sepia officinalis* (Linnaeus, 1758), cultured in the Algarve (South Portugal). *Aquaculture International*, **10**, 207–220
- Domingues, P., Poirier, R., Dickel, L. et al. (2003) Effects of culture density and live prey on growth and survival of juvenile cuttlefish, *Sepia officinalis*. *Aquaculture International*, **11**, 225–242
- Domingues, P., Sykes, A., Sommerfield, A. et al. (2004) Growth and survival of cuttlefish (*Sepia officinalis*) of different ages fed crustaceans and fish. Effects of frozen and live prey. *Aquaculture*, **229**, 239–254
- Domingues, P.M., DiMarco, F.P., Andrade, J.P. et al. (2005) Effect of artificial diets on growth, survival and condition of adult cuttlefish, *Sepia officinalis* Linnaeus, 1758. *Aquaculture International*, **13**, 423–440
- Douglas, R.H., Williamson, R. and Wagner, H.J. (2005) The pupillary response of cephalopods. *Journal of Experimental Biology*, **208**, 261–265
- Dunlop, C. and King, N. (2008) *Cephalopods: Octopuses and Cuttlefish for the Home Aquarium*. tfh Publications, Neptune City
- Ferguson, G.P., Messenger, J.B. and Budelmann, B.U. (1994) Gravity and light influence the countershading reflexes of the cuttlefish *Sepia officinalis*. *Journal of Experimental Biology*, **191**, 247–256
- Fiorito, G. and Scotto, P. (1992) Observational learning in *Octopus vulgaris*. *Science*, **256**, 545–547
- Forsythe, J.W., DeRusha, R.H. and Hanlon, R.T. (1994) Growth, reproduction and life-span of *Sepia officinalis* (Cephalopoda, Mollusca) cultured through seven consecutive generations. *Journal of Zoology*, **233**, 175–192
- Forsythe, J.W., Lee, P.G., Walsh, L.S. et al. (2002) The effects of crowding on growth of the European cuttlefish, *Sepia officinalis* Linnaeus, 1758 reared at two temperatures. *Journal of Experimental Marine Biology and Ecology*, **269**, 173–185
- Gozansky, E.K., Ezell, E.L., Budelmann, B.U. et al. (2003) Magnetic resonance histology: in situ single cell imaging of receptor cells in an invertebrate (*Loliguncula brevis*, Cephalopoda) sense organ. *Magnetic Resonance Imaging*, **21**, 1019–1022
- Guerra, A. (1992) Mollusca, Cephalopoda. In: *Fauna Iberica*, Vol. 1. Ed. Ramos, M.A., pp. 1–327. Museo Nacional de Ciencias Naturales CSIC, Madrid



- Gutfreund, Y., Flash, T., Yarom, Y. *et al.* (1996) Organization of octopus arm movements: a model system for studying the control of flexible arms. *Journal of Neuroscience*, **16**, 7297–7307
- Gutfreund, Y., Flash, T., Fiorito, G. *et al.* (1998) Patterns of arm muscle activation involved in octopus reaching movements. *Journal of Neuroscience*, **18**, 5976–5987
- Hanlon, R.T. and Forsythe, J.W. (1990) Diseases of mollusca: cephalopoda. 1.1 Diseases caused by microorganisms. In: *Diseases of Marine Animals*, Vol. 3. Ed. Kinne, O., pp. 23–46. Biologische Anstalt Helgoland, Hamburg
- Hanlon, R. and Forsythe, J. (2008) Sexual cannibalism by *Octopus cyanea* on a Pacific coral reef. *Marine and Freshwater Behavior and Physiology*, **41**, 19–28
- Hanlon, R.T. and Messenger, J.B. (1996) *Cephalopod Behaviour*. Cambridge University Press, Cambridge
- Hochberg, F.G. (1990) Diseases of mollusca: cephalopoda. 1.2 Diseases caused by protists and metazoans. In: *Diseases of Marine Animals*, Vol. 3. Ed. Kinne, O., pp. 47–227. Biologische Anstalt Helgoland, Hamburg
- Hochner, B., Brown, E., Langella, M. *et al.* (2003) A learning and memory area in the *Octopus* brain manifests a vertebrate-like long-term potentiation. *Journal of Neurophysiology*, **90**, 3547–3554
- Hochner, B., Shomrat, T. and Fiorito, G. (2006) The octopus: a model for a comparative analysis of the evolution of learning and memory. *Biological Bulletin*, **210**, 308–317
- Iglesias, J., Sánchez, F.J., Bersano, J.G.F. *et al.* (2007) Rearing of *Octopus vulgaris* paralarvae: Present status, bottlenecks and trends. *Aquaculture*, **266**, 1–15
- Jereb, P. and Roper, C.F.E. (Eds) (2005) *Cephalopods of the World, an Annotated and illustrated Catalogue of Cephalopod Species Known to Date*, Vol. 1. ttf Publications, Neptune City
- Jereb, P., Ragonese, R. and von Boletzky, S. (1991) *Squid Age Determination using Statoliths*. Note Technique e Reprints dell'Istituto di Tecnologia della Pesca e del Pescato. Special Publication 1. Mazara del Vallo, Italy
- Karson, M.A., Boal, J.G. and Hanlon, R. (2003) Experimental evidence for spatial learning in cuttlefish (*Sepia officinalis*). *Journal of Comparative Psychology*, **117**, 149–155
- Kier, W.M. and Smith, K.K. (1985) Tongues, tentacles and trunks: the biomechanics of movement in muscular-hydrostats. *Zoological Journal of the Linnean Society*, **83**, 307–324
- Komak, S., Boal, J.G., Dickel, L. *et al.* (2005) Behavioural responses of juvenile cuttlefish (*Sepia officinalis*) to local water movements. *Marine and Freshwater Behavior and Physiology*, **38**, 117–125
- Kuba, M.J., Byrne, R.A., Meisel, D.V. *et al.* (2006) When do octopuses play? Effects of repeated testing, object type, age, and food deprivation on object play in *Octopus vulgaris*. *Journal of Comparative Psychology*, **120**, 184–190
- Koueta, N., Boucaud-Camou, E. and Noel, B. (2002) Effect of enriched natural diet on survival and growth of juvenile cuttlefish *Sepia officinalis* L. *Aquaculture*, **203**, 293–310
- Landman, N.H., Davis, R.A. and Mapes, R.H. (Eds) (2007) *Cephalopods Present and Past: New Insights and Fresh Perspectives*. Springer, Heidelberg
- Landman, N.H., Tanabe, K. and Davis, R.A. (Eds) (1996) *Ammonioid Palaeobiology*. Plenum Press, New York
- Lewbart, G.A. (Ed.) (2006) *Invertebrate Medicine*. Blackwell Publishing, Iowa
- Mangold, K. (Ed.) (1989) *Céphalopodes*. In: *Traité de Zoologie*, Vol. 5(4). Ed. Grassé, P.P., pp. 1–804. Masson, Paris
- Mather, J.A. (2001) Animal suffering: An invertebrate perspective. *Journal of Applied Animal Welfare Science*, **4**, 151–156
- Mather, J.A. (2008) Cephalopod consciousness: Behavioural evidence. *Consciousness and Cognition*, **17**, 37–48
- Mather, J.A. and Anderson, R.C. (2007) Ethics and invertebrates: a cephalopod perspective. *Diseases of Aquatic Organisms*, **75**, 119–129
- Mäthger, L.M. and Hanlon, R. (2006) Anatomical basis for camouflaged polarized light communication in squid. *Biological Letters*, **2**, 494–496
- Matzner, H., Gutfreund, Y. and Hochner, B. (2000) The neuromuscular system of the flexible arm of the octopus: physiological characterization. *Journal of Neurophysiology*, **83**, 1315–1328
- Melzner, F., Bock, C. and Pörtner, H.O. (2006) Critical temperatures in the cephalopod *Sepia officinalis* investigated using in vivo <sup>31</sup>P NMR spectroscopy. *Journal of Experimental Biology*, **209**, 891–906
- Melzner, F., Mark, F.C. and Pörtner, H.O. (2007) Role of blood-oxygen transport in thermal tolerance of the cuttlefish, *Sepia officinalis*. *Integrative Comparative Biology*, **47**, 645–655
- Messenger, J.B. (1979) The nervous system of *Loligo*. IV. The peduncle and olfactory lobes. *Philosophical Transactions of the Royal Society London B*, **285**, 275–309
- Moltschaniwskyj, N.A., Hall, K., Lipinski, M.R. *et al.* (2007) Ethical and welfare considerations when using cephalopods as experimental animals. *Reviews in Fish Biology and Fisheries*, **17**, 455–476
- Neumeister, H. and Budelmann, B.U. (1997) Structure and function of the *Nautilus* statocysts. *Philosophical Transactions of the Royal Society London B*, **352**, 1565–1588
- Nixon, M. and Young, J.Z. (2003) *The Brains and Lives of Cephalopods*. Oxford University Press, Oxford
- Norman, M. (2000) *Cephalopods: A World Guide*, 2nd edn. ConchBooks, Hackenheim
- Novicki, A., Messenger, J.B., Budelmann, B.U. *et al.* (1992) (<sup>14</sup>C) Deoxyglucose labelling of functional activity in the cephalopod central nervous system. *Proceedings of the Royal Society of London B*, **249**, 7–82
- Oestmann, D.J., Scimeca, J.M., Forsythe, J.W. *et al.* (1997) Special considerations for keeping cephalopods in laboratory facilities. *Contemporary Topics in Laboratory Animal Science*, **36**, 89–93
- Packard, A. (1972) Cephalopods and fish: the limits of convergence. *Biological Review*, **47**, 241–307
- Packard, A., Karlsen, H.E. and Sand, O. (1990) Low frequency hearing in cephalopods. *Journal of Comparative Physiology A*, **166**, 501–505
- Payne, A.I.L., Lipinski, M.R., Clarke, M.R. *et al.* (Eds) (1998) *Cephalopod: Biodiversity, Ecology and Evolution*. *South African Journal of Marine Science*, **20**, 143–151
- Plän, T. (1987) *Functional neuroanatomy of sensory-motor lobes of the brain of Octopus vulgaris*. PhD thesis, University of Regensburg
- Preuss, T. and Budelmann, B.U. (1995a) A dorsal light reflex in a squid. *Journal of Experimental Biology*, **198**, 1157–1159
- Preuss, T. and Budelmann, B.U. (1995b) Proprioceptive hair cells on the neck of the squid *Lolliguncula brevis*: a sense organ in cephalopods for the control of head-to-body position. *Philosophical Transactions of the Royal Society London B*, **349**, 153–178
- Quast, M.J., Neumeister, H., Ezell, E.L. *et al.* (2001) MR microscopy of cobalt-labeled nerve cells and pathways in an invertebrate brain (*Sepia officinalis*, Cephalopoda). *Magnetic Resonance in Medicine*, **45**, 575–579
- Saidel, W.M., Lettvin, J.Y. and MacNichol, E.F. (1983) Processing of polarized light by squid photoreceptors. *Nature*, **304**, 534–536
- Saidel, W.M., Shashar, N., Schmolesky, M.T. *et al.* (2005) Discriminative responses of squid (*Loligo pealeii*) photoreceptors to polarized light. *Comparative Biochemistry and Physiology*, **142**, 340–346
- Shashar, N., Hanlon, R.T. and Petz, A.D. (1998) Polarization vision helps detect transparent prey. *Nature*, **393**, 222–223
- Shashar, N., Milbury, C.A. and Hanlon, R. (2002) Polarization vision in cephalopods: neuroanatomical and behavioral features that illustrate aspects of form and function. *Marine and Freshwater Behavior and Physiology*, **35**, 57–68
- Sinn, D.L. and Moltschaniwskyj, N.A. (2005) Personality traits in dumpling squid (*Euprymna tasmanica*): context-specific traits and



- their correlation with biological characteristics. *Journal of Comparative Psychology*, **129**, 99–110
- Smith, K.K. and Kier, W.M. (1989) Trunks, tongues and tentacles: moving with skeletons of muscle. *American Scientist*, **77**, 28–35
- Sumbre, G., Gutfreund, Y., Fiorito, G. *et al.* (2001) Control of octopus arm extension by a peripheral motor program. *Science*, **293**, 1845–1848
- Sumbre, G., Fiorito, G., Flash, T. *et al.* (2005) Motor control of the octopus flexible arm. *Nature*, **433**, 595–596
- Sweeney, M.J., Roper, C.F.E., Mangold, K.M. *et al.* (Eds) (1992) 'Larval' and juvenile cephalopods: a manual for their identification. *Smithsonian Contributions to Zoology*, **513**, 1–282
- Sykes, A.V., Domingues, P.M., Correia, M. *et al.* (2006) Cuttlefish culture – state of the art and future trends. *Vie et Milieu*, **56**, 129–137
- Sykes, A.V., Domingues, P.M., Loyd, M. *et al.* (2003) The influence of culture density and enriched environments on the first stage culture of young cuttlefish, *Sepia officinalis* (Linnaeus, 1758). *Aquaculture International*, **11**, 531–544
- Tu, Y. and Budelmann, B.U. (2000) Inhibitory effect of cyclic guanosine 3',5'-monophosphate (cGMP) on the afferent resting activity in the cephalopod statocyst. *Brain Research*, **880**, 65–69
- Vidal, E.A.G., DiMarco, F.P., Wormuth, J.H. *et al.* (2002a) Optimizing rearing conditions of hatchling loliginid squid. *Marine Biology*, **140**, 117–127
- Vidal, E.A.G., DiMarco, F.P., Wormuth, J.H. *et al.* (2002b) Influence of temperature and food availability on survival, growth and yolk utilization in hatchling squid. *Bulletin of Marine Science*, **71**, 915–931
- Villanueva, R. and Bustamente, P. (2006) Composition in essential and non-essential elements of early stages of cephalopods and dietary effects on the elemental profiles of *Octopus vulgaris* paralarvae. *Aquaculture*, **261**, 225–240
- Voight, J.R., Pörtner, H.O. and O'Dor, R.K. (1994) A review of ammonia-mediated buoyancy in squids (Cephalopoda: Teuthoidea). *Marine and Freshwater Behavior and Physiology*, **25**, 193–203
- Voss, N.A., Vecchione, M., Toll, R.B. *et al.* (eds) (1998) Systematics and biogeography of cephalopods. *Smithsonian Contributions to Zoology*, **586**, 1–599
- Walker, I.D., Dawson, D.M., Flash, T. *et al.* (2005) Continuum robot arms inspired by cephalopods. *Proceedings of SPIE*, **5804**, 303–314
- Williamson, J.A., Fenner, P.J., Burnett, J.W. *et al.* (eds) (1996) *Venomous and Poisonous Animals: A Medical and Biological Handbook*. University of New South Wales Press, Sydney
- Williamson, R. (1988) Vibration sensitivity in the statocysts of the Northern octopus, *Eledone cirrosa*. *Journal of Experimental Biology*, **134**, 451–454
- Williamson, R. (1995) The statocysts of cephalopods. In: *Cephalopod Neurobiology*. Eds Abbott, N.J., Williamson, R. and Maddock, L., pp. 503–520. Oxford University Press, Oxford
- Williamson, R. and Budelmann, B.U. (1991) Convergent inputs to octopus oculomotor neurones demonstrated in a brain slice preparation. *Neuroscience Letters*, **121**, 215–218
- Williamson, R. and Chrachri, A. (2004) Cephalopod neural networks. *Neurosignals*, **13**, 87–98
- Yekutieli, Y., Mitelman, R., Hochner, B. *et al.* (2007) Analyzing octopus movements using three dimensional reconstruction. *Journal of Neurophysiology*, **98**, 1775–1790
- Yekutieli, Y., Sagiv-Zohar, R., Aharonov, R. *et al.* (2005a) Dynamic model of the octopus arm. I. Biomechanics of the octopus reaching movement. *Journal of Neurophysiology*, **94**, 1443–1458
- Yekutieli, Y., Sagiv-Zohar, R., Hochner, B. *et al.* (2005b) Dynamic model of the octopus arm. II. Control of reaching movements. *Journal of Neurophysiology*, **94**, 1459–1468
- Young, J.Z. (1965) The central nervous system of *Nautilus*. *Philosophical Transactions of the Royal Society London B*, **249**, 1–25
- Young, J.Z. (1971) *The Anatomy of the Nervous system of Octopus vulgaris*. Clarendon Press, Oxford
- Young, J.Z. (1974) The central nervous system of *Loligo*. I. The optic lobe. *Philosophical Transactions of the Royal Society London B*, **267**, 263–302
- Young, J.Z. (1976) The nervous system of *Loligo*. II. Suboesophageal centres. *Philosophical Transactions of the Royal Society London B*, **274**, 101–167
- Young, J.Z. (1977) The nervous system of *Loligo*. III. Higher motor centres: the basal supraoesophageal lobes. *Philosophical Transactions of the Royal Society London B*, **276**, 351–398
- Young, J.Z. (1979) The nervous system of *Loligo*. V. The vertical lobe complex. *Philosophical Transactions of the Royal Society London B*, **285**, 311–354